Looking Ahead: Applying new Genomic Technologies to Accelerate Genetic Improvement in Beef Cattle

Angela Cánovas¹

Abstract. In recent years, producers have combined the use of phenotypic appraisal and the estimation of breeding values (PTA or EPD) to make genetic selection decisions in beef and dairy cattle that have resulted in a steady genetic gain of 2% per year. However, the most extensive application of genomics has occurred in dairy with the estimation of molecular breeding values that has increased selection efficiency to a much higher order of magnitude. Despite a growing molecular and physiological understanding of complex traits, little is known about the genes determining the traits and their precise function, and a significant unexplained source of variation of phenotypes remains in livestock. Within this context, a more complete understanding of the genes and regulatory pathways and networks involved in economically important traits (i.e. feed efficiency and methane emissions, meat quality and carcass traits) in beef cattle will provide knowledge to help improve genetic selection and reproductive management. Therefore, high throughput -OMICS technology (i.e., transcriptomics, metagenomics, metabolomics, as well as epigenetics and gene networks amongst several others), will complement these tools and further advance identification of functional genes within a systems biology approach.

Key words: Beef cattle, Genomics, -OMICS technologies.

Mirando hacia el futuro: Aplicación de Tecnologías Genómicas Nuevas para Acelerar el Mejoramiento Genético en Ganado de Carne

Resumen. En los últimos años, los productores han combinado el uso de la evaluación fenotípica y la estimación de los valores genéticos (PTA o EPD) para tomar decisiones y seleccionar genéticamente el vacuno de carne y lechero resultando en una ganancia genética constante de 2% por año. Sin embargo, la aplicación de la genómica se ha realizado de modo más extenso en la estimación de los valores genéticos moleculares en el vacuno de leche aumentando considerablemente la eficiencia y precisión de la selección en esta especie. A pesar de que cada vez es mayor el conocimiento molecular y fisiológico de los caracteres complejos, poco se sabe todavía acerca de los genes involucrados así como la función y el papel que desempeñan, lo que conlleva al desconocimiento de un porcentaje significativo de variación que podría explicar fenotipos de importancia económica en vacuno. En este contexto, un mejor conocimiento de los genes y los procesos metabólicos y vías de regulación en los que están involucrados los genes asociados con caracteres de importancia económica en vacuno de carne, como por ejemplo, la eficiencia alimentaria y emisión de metano, calidad de la carne y caracteres relacionados con la canal, proporcionaría el conocimiento para ayudar a mejorar la precisión de la selección genética y el manejo reproductivo. Por lo tanto, las tecnologías de alto rendimiento (llamadas -ómicas) disponibles actualmente en genómica animal, (como por ejemplo la transcriptómica, la metagenómica, la metabolómica así como la epigenética, y las redes génicas asociadas, entre otras) combinadas con la metodología de biología de sistemas, ayudaran a la identificación de genes funcionales que desempeñan un papel clave en la regulación de los caracteres de interés económico.

Palabras clave: Genómica, Tecnologías –ÓMICAS, Vacuno de carne.

Background

The field of genetics and genomics in most of the livestock species has experienced a dramatic technological revolution in the last 5 years. In the

earlier 2000's, the emphasis in livestock genetics was in linkage maps with single markers and quantification of some genes using real time qPCR. The first bovine genome assembly was published in 2009 (Elsik *et al.*, 2009), and since that time, the development and use of

¹ Centre for Genetic Improvement of Livestock, Department of Animal Bioscience, University of Guelph, 50 Stone Road E, Guelph, ON N1G 2W1, Canada. Email acanovas@uoguelph.ca

various whole genome-omics tools has accelerated research in cattle genetics (Reverter *et al.*, 2013; Snelling *et al.*, 2013). After the development of the livestock genome sequences, new technologies, such as microarrays, were introduced allowing the study of gene expression of the entire transcriptome. With that, the concept of Genetical Genomics was developed with the idea of integrating structural and functional genomic data, combining gene expression from

microarray technology with genotype data from marker genotypes. Now, the field of Genetical Genomics or the integration of structural and functional genomic data, has expanded as high throughput tools have become available for genomic analysis such as high-density (HD) genotyping-chips (Illumina, San Diego, CA), whole genome sequencing and genotyping by sequencing and RNA-Sequencing to measure the gene expression in the entire transcriptome (in a more accurate way than microarray technology; Wickramasinghe *et al.* 2014) (Figure 1).



Single Genes > Genetical Genomics > System Biology

Figure 1. Evolution in the field of genetics and genomics in most of the livestock species in the last decade. New Genomic technologies and high throughput –OMICS technologies to accelerate genetic improvement in beef cattle.

Currently, with all the new available technologies in livestock combined with statistical methodologies has allowed the integration of structural and functional genomics information with other –OMICS into a systems biology approach to develop a better biological understanding of phenotypes. As part of the genomics tool box the HD-genotyping chips such as 50K and 800K and also whole genome sequencing are now available in most of the livestock species and have been extensively utilized, such as in dairy cattle genetic improvement.

As a part of high throughput tools available for genomic analysis, RNA-Sequencing allows measuring not only gene expression, but also examining genome structure identifying SNP and other structural variation such as insertions, deletions and splice variants (Wickramasinghe *et al.* 2014; Cánovas *et al.* 2010). The expectation is that the integration of all these genomic data will accelerate the genetic improvement by improving accuracy of selection and reducing the generation interval (Figure 1).

Combining the information from the -OMICS technologies together with metabolic pathways and functional/biological analysis into a Systems biology approach allows the identification of functional SNP increasing the accuracy of selection. Trancriptomics using RNA-Sequencing technology measures gene expression of the entire transcriptome, as well as structural variation (i.e. SNP, insertions, deletions and splice variants in coding regions). Also, it allows the identification of the differential expressed splice variants affecting genes overlapping significant SNPs in significant QTL regions. Metagenomics, identifies and quantifies the microbial profiles and determine their functionality in specific bovine samples (i.e. rumen fluid) using 16SrRNA based profiling of microbial populations as well as partial protozoa and fungal 18S rRNA (Handerson et al., 2015). Metabolomics, identifies circulating plasma and/or rumen fluid metabolites as potential biomarkers to assess physiological and hormonal states relative to performance by Gas-chromatography-Massspectrometry (GC-MS) and NMR analysis. For each analysis, an internal standards are included for normalization to allow comparisons between samples. Temporal and treatment changes in metabolites can be determined to be significant after normalization and can be correlated to the microbial profiles and gene expression obtained from RNA-Sequencing. The microbial profiles and/or microbial activities of each

animal can be then defined and associated with the phenotype of interest using a multivariate approach.

The particular benefit of new integrated high throughput genomics technologies will be most likely for genetic improvement of those traits that are difficult to measure such as feed efficiency, methane emission and fertility and reproduction traits.

Beef Cattle Genetic Improvement Using Genomic Technologies

In general, the beef industry is highly segmented (seedstock, cow-calf, feedlot, processor, retailer, and consumer). This segmentation is seen in the way various stakeholders along the supply chain make logical decisions for their own business yet in the light of the entire chain, these decisions are detrimental to the end product and consumer satisfaction. With little or no market signal back along the supply chain, needed changes are not made for the end product to be desired and competitive with other protein choices facing the consumer. Poor information flow along the beef supply chain is leading to massive inefficiency as demonstrated in the dramatic decline in high yielding carcasses in the past decade (down from 66% to 41%). This drop resulted from overfeeding in an attempt to increase the level of intramuscular fat (marbling). The past 30 years has seen a dramatic and consistent decline in per capita beef consumption, due in part as a result of inconsistent product, primarily in the important trait of tenderness. Over the past 30 years (and even more drastically the last 10 years), beef consumption per capita in Canada has declined from 27 to 19 kg, while that of chicken has risen from 21 to 33 kg.

The many studies of the beef industry completed in the past 20 years all identify lack of information flow as a serious impediment to the industry. This is more currently demonstrated by the inclusion of connectedness (information flow along and back the supply chain) as one of four target areas of focus in the Canadian Cattlemen's Association's (CCA's) national beef strategy (beefstrategy.com).

The utilization of genomics in the beef industry has been limited (compare to dairy industry), due in part to the 1) complicating factor of multiple breeds and correspondence of SNP effects, 2) in general, the genetic evaluations are carried out by the multiple breed associations using different methods to calculate the EPDs, 3) reporting different EPDs for different breeds being more difficult the data comparison from different evaluations (Van Eenennaam 2016). International Genetics Solutions (IGS) is trying to leverage a multibreed database that enables the comparison of EPDs across breeds associations (Chianina, Gelbvieh, Limousin, Maine-Anjou, Red Angus, Simmental, Shorthorn, Canadian Simmental, Canadian Gelbvieh, Canadian Limousin, Canadin Angus and Canadian Shorthom).

Genomic approaches offer an opportunity to accelerate the genetic improvement, but genomics requires accurate phenotypes (and genotypes) from genomically linked individual animals. Therefore the focus is on collecting data to enable generation of GEBV for all animals in a population. Industry breeding strategies will then incorporate these new genetic evaluations to improve production efficiency and to provide a more consistent and competitive product to consumers.

Also, beef producers often make breeding decisions without current knowledge of the science, existing systems and tools that they can use on farm to benefit from this research. Taken in isolation, a research finding can be confusing and lack any meaningful connection to the real world of production in which producers make decisions. Thus, producers require more concise, layman's terms presentation of the benefits of implementing change in their businesses, as well as a more clearly defined stepwise approach they can take to implement that change. Therefore, is important to do an effort to translate and transfer the research knowledge associated to genetics and the importance of collecting phenotypes (and genotypes) which is key to affecting change necessary to enhance and accelerate innovation in beef industry.

Using genomics-based approaches to improve beef genetics will result in increased production efficiency, making beef a more attractive protein option than current relative to poultry and pork. Effective use of genomics will also provide the consumer with a more consistently tender product and therefore help to address declining consumption.

Therefore it's important to capture and make use of data in the development of genomically enhanced Expected Progeny Differences (GEPD's) which are simply more accurate indicators of an animal's true genetic merit. Although there are phenotype (related to meat quality and carcass traits) and genotype data collected, more phenotype and genotype data are required to enable generation of GEPDs to incorporate into beef industry breeding strategies. New sets of data will be crucial to enlarge the reference set of animals with genotype and phenotype, but most important, will provide the researchers with additional information collected at the whole chain scope, which will be unique to improve the quality of prediction equations and at the same time, enhance economical analysis of the desired traits.

GEPDs have a significant role to play in the effective selection for these traits. The selection efficiency with GEPDs will be higher with the knowledge gained from functional genomics studies. Despite a growing molecular and physiological understanding of complex traits such as carcass traits, little is known about the genes determining the traits associated to tenderness and their precise function. and a significant unexplained source of variation of phenotypes remains in beef. Within this context, a more complete understanding of the genes and regulatory pathways and networks involved in economically important traits such as tenderness and carcass traits in beef cattle will provide knowledge to help improve genetic selection. By leveraging -OMICS and systems biology, we can develop more robust approaches for genetic selection that can be incorporated to industry breeding plans for improving meat quality focusing on tenderness and carcass traits in beef cattle.

Large databases containing genomic and phenomic information, together with the data generated using the new genomic technologies (for example, gene expression data from RNA-Sequencing together with the results from GWAS (using the phenotypes and genotypes) could be subjected to genome wide association studies to identify quantitative trait loci (QTLs) for carcass quality which will assist in the ranking of functional variants. These analyses can be used to develop prediction equations for gEPDs which are incorporated into multi-trait value indices that perform well in commercial cattle.

Gains in prediction accuracy achievable by the addition of subsets of functional variants to existing panels can be quantified by imputing the variants into training and testing populations already genotyped. Information about variant function can be incorporated into genomic predictions through the use of the BayesRC method (MacLeod *et al.* 2014) or weighted genomic best linear unbiased prediction (GBLUP).

Another important aspect to successfully exploit the power of applying the new genomic technologies to accelerate the genetic improvement in beef cattle is the annotation of the bovine genome. Although the first bovine genome assembly in 2009 and the subsequent releases in the last years, still there are several gaps and errors in the annotation of the current bovine genome. In order to improve the annotation and taking advantage of the results generated using the new technologies available. OMICS there is the international initiative on the Functional Annotation of Animal Genomes (FAANG); a coordinated international action to accelerate Genome to Phenome (see web site for more detail: http://www.faang.org/index). The project is expecting to close the genotype-to-phenotype gap providing new information on genetic variants that explain variation in the target traits that can be used to increase the accuracy of the genomic predictions.

Fertility and Reproduction

In relation to reproduction, an example of combining the new genomic technologies is the analysis of puberty in beef cattle by examining the genes and regulatory pathways and networks involved in this complex physiological event. Puberty is the process by which animals mature into an adult capable of sexual reproduction (Dorn and Biro 2011).

The process to achieve the puberty is similar in the two bovine sub-species (i.e., *Bos indicus* and *Bos taurus*), but occurrs at markedly older ages in *Bos indicus* heifers (Rodrigues *et al.*, 2002; Nogueira, 2004). However, despite a growing molecular and physiological understanding of the reproductive system, knowledge of the precise mechanisms regulating puberty in cattle is limited, and phenotypic identification of animals that undergo puberty at an early age is costly and labor-intensive. Therefore, enhancing our understanding of the genes and regulatory pathways and networks involved in bovine puberty can provide a window to help improve genetic selection and reproductive management in cattle.

Whole genome single nucleotide polymorphism (SNP)-chip and RNA-Sequencing data from the hypothalamus have been used to construct gene networks associated with puberty in cattle (Fortes *et al.* 2010 and 2011). Results from these approaches allowed postulating that regulatory loci underlying the quantitative trait loci (QTL) associated with heifer fertility traits influence puberty. Livestock production

traits are usually complex and involve multiple tissues. Therefore, the transcriptome of five tissues related to reproduction (i.e. hypothalamus, pituitary gland, ovary, uterus, and endometrium) has been characterized as well as tissues known to be relevant to growth and metabolism needed for cattle to achieve puberty (i.e., longissimus dorsi muscle, fat, and liver) in PRE and puberty heifers usina POST **RNA-Sequencing** (Cánovas et al. 2014). In order to exploit the power of complementary -OMICS analyses, PRE and POST pubertv co-expression gene networks were constructed by combining the results from RNA-Sequencing, genome-wide association study (GWAS), and bovine transcription factors. As a result of combining the power of the different genomic technologies this reduced the complexity of the large lists of SNP and/or genes identified associated with puberty (Figure 2). Thus, combining the results from RNA-Sequencing and GWAS identified a total of 25 eQTL associated to heifer fertility. Applying the new genomic technologies and integrating the structural functional genomic data revealed and kev transcriptional regulators (i.e., PITX2, FOXA1, TSG1D1, DACH2, LHX4, PROP1 and SIX6) (Table 1). As a validation of the approach used combining data from several genomic technologies, six genes captured in the cattle network were concordant with the human network that reported 30 loci for age at menarche using other functional analysis (Elks et al., 2010). Results from multiples sources of -omics data will facilitate the design of breeding strategies to improve fertility in Bos indicus-influenced composite cattle (Cánovas et al. 2014). Multi-tissue omics analyses improve understanding of the number of genes and their complex interactions for puberty in cattle. These results also help discovering genes that contain biologically relevant SNP-genotypes to validate in an independent population that can be used in genetic improvement processes of Bos indicus-influenced composite cattle.

Feed Efficiency and Methane Emissions

Feed costs are currently the largest expense in milk and meat production in livestock (approx. 70% of total cost). Therefore, improving feed conversion efficiency will have a direct economic benefit to the dairy and beef industries. In addition to high feed costs, the industry is also under increased pressure to reduce the environmental impact of cattle production, primarily the contribution to greenhouse gases. Genomics-

Ceiba

based approaches can provide an added advantage above traditional methods of genetic improvement as demonstrated for dairy cattle. New sequencing and high-throughput technologies provide new opportunities to further increase selection efficiency and address new traits of importance for consumers (such as environmental sustainability). High-throughput -OMICS technologies including transcriptomics, metagenomics and metabolomics can be combined with existing phenotype/genotype data and information within a systems biology approach. The result is a more complete understanding of the genes, and regulatory pathways and networks involved in economically important traits such as feed efficiency, and integrate this comprehensive information for use in genetic improvement programs.



Figure 2. Combining results from different genomic technologies to identify the key regulator genes and/or functional SNP associated with puberty using the list of: 1) Differentially expressed genes between pre- and post-puberty, 2) Genes tissue specific, 3) Transcription factors and 4) Genes harboring SNP observed with GWAS and associated with indicator traits of puberty (age at puberty as measured by the presence of the first corpus luteum, first service conception and heifer pregnancy).

The goal of including genomic information in genetic evaluation is to increase the accuracy of predicting genetic merit for breeding animals. For breeders to make the best use of genomic data, it needs to be combined with traditional sources of information (i.e. phenotypes and pedigrees) into traditional genetic evaluations. The main advantage of including genomic information in the estimation of breeding values (PTA or EPD) is that it improves the accuracy of EPDs on young animals where there is little other information upon which to base genetic merit estimates. This increase in accuracy is dependent upon how much of the genetic variation is explained by the genomic information.

Genomic approaches offer an opportunity to accelerate the genetic improvement, but genomics requires accurate phenotypes (and genotypes) from genetically-representative individual animals. Therefore the focus is on collecting data to enable generation of GEBV (Genomic Estimated Breeding Values) for all animals in a population. Industry breeding strategies will then incorporate these new genetic evaluations to improve production efficiency and to provide a more consistent and competitive product to consumers.

Using genomics-based approaches to improve dairy genetics will result in increased production efficiency and reduced environmental footprint. Since feed is currently the largest expense in milk production (MacDonald et al. 2007), improving cow efficiency in converting feed to milk will be an economic benefit to the industry. More efficient animals also produce less manure and emit less methane, reducing the industry's environmental footprint. Preliminary estimates show that breeding for increased feed efficiency and reduced methane emission can reduce feed cost in dairy cattle by \$108/cow/year (M. Coffey, unpublished data) and decrease methane emission by an estimated 11-26% (De Haas et al. 2011). In Canada, genetic progress has more than doubled since the implementation of genomics, yielding an additional \$210M/year to the Canadian Dairy Industry (Dr Filippo Miglior, Canadian Dairy Network, 2016).

Genetic advancement in the beef industry has been limited in a number of ways including the high cost of determining genetic merit (low selection efficiency). While genomics has proven highly effective in the dairy industry, the challenges faced by the beef sector will require a more sophisticated approach than what is currently available. Utilization of genomics in the beef industry has been low to date, due in part to the complicating factor of multiple breeds and correspondence of SNP effects. The incorporation of new –OMICS technologies will enhance the accuracy of determination of genetic merit and make genomics more applicable and useful to beef breeders.

Cánovas: Looking Ahead: Applying new Genomic Technologies to Accelerate Genetic Improvement in Beef Cattle

In this way, the new high throughput tools available for genomic analysis will build upon all of the genomics work to date and have the effect of making genomics applicable to an entire sector, which would have a particular benefit to beef production. That paradigm shift will provide beef breeders with a tool that will let them become much more competitive with other protein producing sectors through improved feed efficiency, weight gain in a reduced time span, more consistent product and reduced negative environmental impact.

Table 1. Identification of 25 eQTL associated to heifer fertility and the key regulator genes / functional SNP combining the results from RNA-Sequencing and GWAS.

RNA - Sequencing Analysis						SNP Association Analysis (GWAS)			
Cana	DE	те	Tiesue	Fold	CND	Chr	Desition (hp)	Troit	
Gene		13	IISSUE	Change	5NP	Chr.		Trait	P- value
DPP A 4	1	0	nypothalamus	-4.82	SS61475098	1	54,523,895	HPG	0.00554
TP63	1	1	adipose	5.06	ss86286585	1	78,088,393	FSC	0.00745
INHA	0	1	ovary	-1.26	BovineHD0200031132	2	108,231,291	ACL	0.00504
IL22RA1	1	0	endometrium	-16.78	rs29019426	2	129,348,719	FSC	0.00205
RHEBL1	1	0	pituitary gland	-3.97	ss86320097	5	30,912,029	HPG	0.00372
LYSB	0	1	endometrium	-2.02	BovineHD0500012799	5	44,556,197	ACL	0.00517
ADH6	1	0	endometrium	25	ss61513484	6	26,799,276	HPG	0.00615
MEPE	1	1	pituitary gland	4.71	ss117968512	6	38,286,952	HPG	0.0059
TECRL	0	1	LD muscle	-1.22	BovineHD0600022520	6	81,577,343	ACL	0.00746
LOC777593	1	0	endometrium	15.96	ss86282432	7	61,683,533	HPG	0.00549
MGC157266	1	0	endometrium	-3.76	BovineHD0800018795	8	62,927,382	ACL	0.00715
C10H11ORF46	1	1	hypothalamus	-4.16	BovineHD1000001491	10	4,882,326	ACL	0.00292
NRXN3	1	0	endometrium	-3.92	rs43651752	10	91,751,584	HPG	0.00283
TSHR	1	0	endometrium	4.55	ss117964119	10	93,528,030	HPG	0.00369
NEBL	1	0	endometrium	5.63	ss61500113	13	22,476,667	FSC	0.00157
MOS	0	1	ovary	-2.05	BovineHD1400007249	14	24,973,324	ACL	0.00017
PENK	1	0	pituitary gland	4.54	BovineHD1400007306	14	25,218,861	ACL	0.00061
ELF5	1	0	uterus	-17.19	BovineHD1500018865	15	65,840,403	ACL	0.00879
POU4F2	0	1	endometrium	-2.45	rs41838669	17	11,828,433	HPG	0.00707
FAM19A4	1	0	liver	-4.46	BovineHD2200009370	22	32,820,574	ACL	0.00712
ITIH1	1	0	ovary	3.48	ss86298585	22	48,663,639	HPG	0.00567
CPNE5	1	0	endometrium	-4.95	BovineHD2300002612	23	10,690,330	ACL	0.00743
MMD2	1	0	uterus	19.89	BovineHD2500011055	25	39,622,260	ACL	0.00493
DKK1	1	0	uterus	11.65	rs29024937	26	6,860,529	FSC	0.00817
SYCE1	0	1	liver	1.02	ss61529911	26	50,517,416	HPG	0.00817

In beef cattle, the best genomics prediction using SNP markers has an accuracy of only 20%, which is insufficient for impact and adoption. However, as described by Karisa et al. (2014), metabolites could explain over 75% of the variation in residual feed intake (RFI) and result in ~80% accuracy of prediction. One of the objectives is to reduce the complexity of the large lists of SNP and/or genes associated with a trait of interest to identify the key regulator genes and functional SNP (and genomic regions) associated to feed efficiency and methane emission by integrating aenomics with the -OMICS functional new technologies into a system biology approach for more reliable results that could be implemented in dairy and beef breeding strategies to improve the accuracy of selection. To have a more complete understanding of the biological knowledge and the genes and mutations affecting feed efficiency, it is also important to study the microbiota profile associated to the genotype of the animal and its interactions affecting the feed efficiency in rumen fluid. A metatranscriptome (using also RNA-Sequencing) approach can used to identify the active microbial phylotypes including bacteria, archaea, eukyrotic (fungi and protozoa) and rumen microbial activities from animals with different RFI ranking. Metatranscriptomic can not only generate taxonomy profiles for active organisms, but also can be used to evaluate the activities of the rumen microbiome. Also, metagenomics to identify and quantify microbial profiles and determine their functional capacity in bovine rumen fluid samples. Total DNA is extracted and partial bacterial and archaeal 16S rRNA gene as well as partial protozoa and fungal 18S rRNA gene is amplified as previously described by Handerson et al. (2015) and sequenced using Illumina-Miseq. The purpose of this analysis is to profile the rumen microbiota and rumen microbial metagenomic profiles generated can be correlated with feed efficiency and methane emission phenotypes.

References

- Cánovas, A., G. Rincon, A. Islas-Trejo, S. Wickramasinghe, and J.F. Medrano. 2010. SNP discovery in the bovine milk transcriptome using RNA-Seq technology. Mammalian Genome 21: 592–598.
- Cánovas, A., A. Reverter, K. DeAtley, R. Ashley, M. Colgrave, M. Fortes, A. Islas-Trejo, S. Lehnert, L. Porto-Neto, G. Rincón, G. Silver, W. Snelling, J.F. Medrano, and M.G Thomas. 2014. Multi Tissue Omics Analyses Reveals Molecular Regulatory Networks for Puberty in Composite Beef Cattle. PLOS One 9(7):e102551.

- de Haas, Y., J.J. Windig, M.P.L. Calus, J. Dijkstra, M. de Haan, A. Bannink, and R.F. Veerkamp. 2011. Genetic parameters for predicted methane production and potential for reducing enteric emissions through genomic selection. Journal of Dairy Science 94:6122– 6134.
- Dorn, L.D., and F.M. Biro. 2011. Puberty and its measurement: a decade in review. Journal of Research Adolescence 21: 180–195.
- Elsik, C.G., R.L. Tellam, and K.C. Worley. 2009. The genome sequence of tuarine cattle: a window to ruminant biology and evolution. Science 324: 522–528.
- Elks, C.E., J.R. Perry, P. Sulem, D.I. Chasman, N. Franceschini, C. He, K.L. Lunetta, J.A. Visser, E.M. Byrne, D.L. Cousminer, D.F. Gudbjartsson, T. Esko, B. Feenstra, J.J. Hottenga, D.L. Koller, Z. Kutalik, P. Lin, M. Mangino, M. Marongiu, P.F. McArdle, A.V. Smith, L. Stolk, S.H. van Wingerden, J.H. Zhao, E. Albrecht, T. Corre, E. Ingelsson, C. Havward, P.K. Magnusson, E.N. Smith, S. Ulivi, N.M. Warrington, L. Zgaga, H. Alavere, N. Amin, T. Aspelund, S. Bandinelli, I. Barroso, G.S. Berenson, S. Bergmann, H. Blackburn, E. Boerwinkle, J.E. Buring, F. Busonero, H. Campbell, S.J. Chanock, W. Chen, M.C. Cornelis, D. Couper, A.D. Coviello, P. d'Adamo, U. de Faire, E.J. de Geus, P. Deloukas, A. Döring, G.D. Smith, D.F. Easton, G. Eiriksdottir, V. Emilsson, J. Eriksson, L. Ferrucci, A.R. Folsom, T. Foroud, M. Garcia, P. Gasparini, F. Geller, C. Gieger; Giant Consortium, V. Gudnason, P. Hall, S.E. Hankinson, L. Ferreli, A.C. Heath, D.G. Hernandez, A. Hofman, F.B. Hu, T. Illig, M.R. Järvelin, A.D. Johnson, D. Karasik, K.T. Khaw, D.P. Kiel, T.O. Kilpeläinen, I. Kolcic, P. Kraft, L.J. Launer, J.S. Laven, S. Li, J. Liu, D. Levy, N.G. Martin, W.L. McArdle, M. Melbye, V. Mooser, J.C. Murray, S.S. Murray, M.A. Nalls, P. Navarro, M. Nelis, A.R. Ness, K. Northstone, B.A. Oostra, M. Peacock, L.J. Palmer, A. Palotie, G. Paré, A.N. Parker, N.L. Pedersen, L. Peltonen, C.E. Pennell, P. Pharoah, O. Polasek, A.S. Plump, A. Pouta, E. Porcu, T. Rafnar, J.P. Rice, S.M. Ring, F. Rivadeneira, I. Rudan, C. Sala, V. Salomaa, S. Sanna, D. Schlessinger, N.J. Schork, A. Scuteri, A.V. Segrè, A.R. Shuldiner, N. Soranzo, U. Sovio, S.R. Srinivasan, D.P. Strachan, M.L. Tammesoo, E. Tikkanen, D. Toniolo, K. Tsui, L. Tryggvadottir, J. Tyrer, M. Uda, R.M. van Dam, J.B. van Meurs, P. Vollenweider, G. Waeber, N.J. Wareham, D.M. Waterworth, M.N. Weedon, H.E. Wichmann, G. Willemsen, J.F. Wilson, A.F. Wright, L. Young, G. Zhai, W.V. Zhuang, L.J. Bierut, D.I. Boomsma, H.A. Boyd, L. Crisponi, E.W. Demerath, C.M. van Duijn, M.J. Econs, T.B. Harris, D.J. Hunter, R.J. Loos, A. Metspalu, G.W. Montgomery, P.M. Ridker, T.D. Spector, E.A. Streeten, K. Stefansson, U. Thorsteinsdottir, A.G. Uitterlinden, E. Widen, J.M. Murabito, K.K. Ong, and A. Murrav, 2010. Thirty new loci for age at menarche identified by a metaanalysis of genome-wide association studies. Nature

Cánovas: Looking Ahead: Applying new Genomic Technologies to Accelerate Genetic Improvement in Beef Cattle

Genetics 42: 1077–1085.

- Fortes, M.R.S., A. Reverter, Y. Zhang, E. Collis, S.H. Nagaraj, N.N. Jonsson, K.C. Prayaga, W. Barris, and R.J. Hawken. 2010. Association weight matrix for the genetic dissection of puberty in beef cattle. Proceedings National Academic of Science 107: 13642–13647.
- Fortes, M.R.S., A. Reverter, S.H. Nagaraj, Y. Zhang, N.N. Jonsson, W. Barris, S. Lehnert, G.B. Boe-Hansen, and R.J. Hawken. 2011. A single nucleotide polymorphismderived regulatory gene network underlying puberty in 2 tropical breeds of beef cattle. Journal of Animal Science 89: 1669–1683.
- Henderson, G., F. Cox, S. Ganesh, A. Jonker, W. Young, and P.H. Janssen. (Global Rumen Census Collaborators). 2015. Rumen microbial community composition varies with diet and host, but a core microbiome is found across a wide geographical range. Scientific Report 25: 14567.
- MacDonald, J.M., E.J. O'Donoghue, W.E. McBride, R.F. Nehring, C.L. Sandretto, and R. Mosheim. 2007. Profits, Costs, and the Changing Structure of Dairy Farming. USDA.
- MacLeod, H.D., H.D. Daetwyler, P.J. Bowman, A.J. Chamberlian, C.J. Vander Jagt, A. Capitan, H. Pausch, P. Stothard, X. Liao, C. Schrooten, E. Mullaart, R. Fries, B. Guldbrandtsen, M.S. Lund, D.A. Boichard, R.F. Veerkamp, C.P. VanTassell, B. Gredler, T. Druet, A. Bagnato, J. Vilkki, D.J. deKoning, E. Santus, and M.E.Goddard. 2014. Genomic Prediction from Whole Genome Sequence in Livestock: the 1000 Bull Genomes Project. Proceedings, 10th World Congress of Genetics Applied to Livestock Production.

- Nogueira, G.P. 2004. Puberty in South American Bos indicus (Zebu) cattle. Animal Reproduction Science 82– 83: 361–372.
- Reverter, A., and M.R.S. Fortes. 2013. Breeding and Genetics Symposium: building single nucleotide polymorphism-derived gene regulatory networks: toward functional genome wide association studies. Journal of Animal Science 91: 530–536.
- Rodrigues, H.D., J.E. Kinder, and L.A. Fitzpatrick. 2002. Estradiol regulation of luteinizing hormone secretion in heifers of two breed types that reach puberty at different ages. Biology Reproduction 66: 603–609.
- Snelling, W.M., R.A. Cushman, J.W. Keele, C. Maltecca, M.G. Thomas, M.R. Fortes, and A. Reverter. 2013. Breeding and Genetics Symposium: networks and pathways to guide genomic selection. Journal of Animal Science 91: 537–552.
- Van Eenennaam, A. 2016. Recent Developments in Genetic Evaluations and Genomic Testing. www.ebeef.org.
- Wickramasinghe, S., A. Cánovas, G. Rincon, and J.F. Medrano. 2014. Review: RNA-Seq applications in Livestock. Livestock (Review) 166, 206-216.

Received for publication on April 1, 2016. Acepted for publication on July 16, 2016.